



Patient: Jane A. Doe
DOB/Gender: 10/10/61 (58 yrs) - Female
Patient ID/MRN: 123456
Date Collected: 04/09/2019



Case#/Status: X19-00352 - Final
Report Category:
Neoplastic



Provider: John Doe, M.D.
Hematology Oncology Associates
Tel: 800-123-4567
Fax: 800-765-4321



DIAGNOSIS:

Lymphoplasmacytic lymphoma (see comment)



COMMENT

The overall findings are consistent with involvement by a LOW GRADE NON-HODGKIN B-CELL LYMPHOMA. Although the plasma cell component is morphologically limited and a clonal population of plasma cells was not definitively detected by flow cytometry or immunohistochemistry, in the setting of a monoclonal IgM of approximately 2 g/dL, the presence of a MYD88 mutation is most consistent with involvement by a LYMPHOPLASMACYTIC LYMPHOMA. Correlation with clinical and laboratory findings is advised.

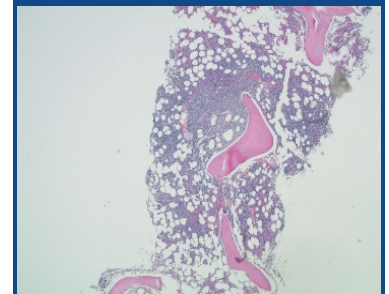
All myeloid and lymphoid neoplasms are now classified and named in accordance with the newly revised 2017 version of the WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues.



COMPONENT DIAGNOSES

- Biopsy:** Non-Hodgkin B-cell lymphoma (see comment)
- Aspirate:** Maturing trilineage hematopoiesis with frequent small lymphocytes
- Flow Cytometry:** Suspicious for a non-Hodgkin B-cell lymphoproliferative disorder (see comment)
- Karyotyping:** Normal female karyotype
- FISH:** No Clonal Abnormalities Detected with probes specific for recurrent abnormalities in Plasma Cell Myeloma (see interpretation for probes tested)
- Molecular:** MYD88 mutation(s) detected: P.L265P (see comment)

Lymphoid Infiltrate



CLINICAL DATA

Monoclonal paraproteinemia.

Received CBC, reported on 4/9/2019: WBC 6.3; RBC 4.36; HGB 12.9; HCT 39.6; MCV 91; MCH 29.7; MCHC 32.6; RDW 14.4; PLT 213; MPV 7.1; LYM 37.4%; MON 10.1%; NEU 'NP'; EOS 'NP'; BAS 'NP' (NP = not provided)

Electronically Signed By: Frank Bauer, MD (04/18/19 17:39)

DIAGNOSIS:

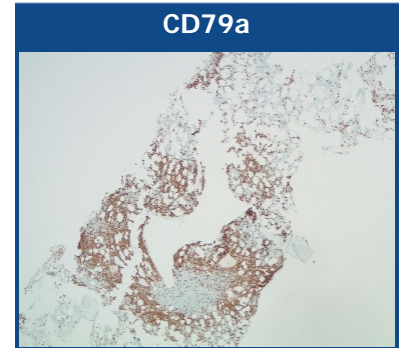
Bone marrow, core & clot biopsies: non-Hodgkin B-cell lymphoma (see comment)

COMMENT

The findings are consistent with involvement by a NON-HODGKIN B-CELL LYMPHOMA with a differential that includes a marginal zone lymphoma and lymphoplasmacytic lymphoma. Correlation with clinical and laboratory findings is advised.

MICROSCOPIC DESCRIPTION

Marrow Cellularity: Mildly hypercellular (60%)
 Infiltrate: Approximately 30% of the cellularity is comprised of an interstitial infiltrate of lymphocytes that are small sized with round nuclei, condensed chromatin, indistinct nucleoli and small amounts of cytoplasm occurring singly and in one large aggregate.
 Myeloid Maturation: Normal
 Erythroid Maturation: Normal
 Myeloid/Erythroid Ratio: Mildly increased
 Megakaryocytes: Normal in number and morphology
 Granulomas: Not identified
 Iron Stain: No stainable iron is seen in this decalcified specimen
 Marrow Reticulin: Mild increase in fibrosis is noted in association with the lymphoid aggregate
 Marrow Trabeculae: Normal
 Clot preparation: Similar findings to the core biopsy
 PAS / Giemsa: Examined
 Special Stains: Giemsa, Iron, PAS, Reticulin
 Immunostains: CD20, CD79a and Pax-5 highlight numerous lymphocytes. An aggregate of CD3 positive T cells is seen. CD138 positive plasma cells account for less than 5% of the marrow cellularity and are polyclonal for immunoglobulin kappa and lambda light chains.



Additional Studies:

Stain	Result
CD3 (MRQ-39)	See microscopic description above
CD20 (L26)	See microscopic description above
CD79a (SP18)	See microscopic description above
CD138/syndecan-1 (B-A38)	See microscopic description above
Kappa (L1C1)	See microscopic description above
Lambda (Lamb14)	See microscopic description above
PAX5 (SP34)	See microscopic description above

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GROSS DESCRIPTION:

- The specimen is received in formalin labeled with the patient's initials and requisition number, and consists of 1 piece of bone marrow core measuring 1.0 x 0.2 x 0.2 cm. The specimen is submitted in 1 cassette after decalcification.
- The specimen is received in formalin labeled with patient's initials and requisition number, and consists of 1 piece of bone marrow clot measuring 0.5 x 1.4 x 1.4 cm. The specimen is submitted in 1 cassette.

Disclaimer: The adequacy of staining is verified by the appropriate positive and negative controls. The reagents used for these assays are analyte specific reagents (ASR). Their performance characteristics have been validated by Precipio, Inc., New Haven, CT. They have not been reviewed by the FDA. The FDA has deemed that such approval is unwarranted. These assays are for clinical use and should not be viewed as experimental or "research use only".

Patient: Jane A. Doe

Case #: X19-00352

DIAGNOSIS:

Bone marrow, aspirate: Maturing trilineage hematopoiesis with frequent small lymphocytes

SMEAR REVIEW

The marrow aspirate smear is spicular and cellular with maturing trilineage hematopoiesis and scattered small lymphocytes. Megakaryocytes are normal in number and morphology. The myeloid : erythroid (M:E) ratio is approximately 8:1. Erythroid maturation is normal. Myeloid maturation is normal. No increase in immature cells is noted. Scant, focal iron is seen without ring sideroblasts on iron stain of the marrow aspirate.

Number of cells counted: 222

Cell Type	Percent	Ref. Range
Blasts	0 % ↓	0.3 - 3.0 %
Immature myeloid	9 % ↓	12.0 - 21.0 %
Mature myeloid	63 % ↑	35.0 - 55.0 %
Eosinophils	3 %	1.0 - 3.0 %
Basophils	0 %	0.0 - 1.0 %
Lymphocytes	16 % ↑	10.0 - 15.0 %
Plasma cells	1 %	0.0 - 1.0 %
Monocytes	0 %	0.0 - 1.0 %
Erythroid	8 % ↓	15.0 - 25.0 %
M:E ratio	8:1 ↑	2 - 4:1

Electronically Signed By: Frank Bauer, MD (04/12/19 17:37)

DIAGNOSIS:

Bone marrow, aspirate: Suspicious for a non-Hodgkin B-cell lymphoproliferative disorder (see comment)

COMMENT

The differential diagnosis includes lymphoplasmacytic lymphoma and a marginal zone lymphoma. Most cases of chronic lymphocytic lymphoma, mantle cell lymphoma and follicular lymphoma will express either CD5 or CD10. Correlation with the concurrent bone marrow core and aspirate morphology and cytogenetic findings is advised.

INTERPRETATION

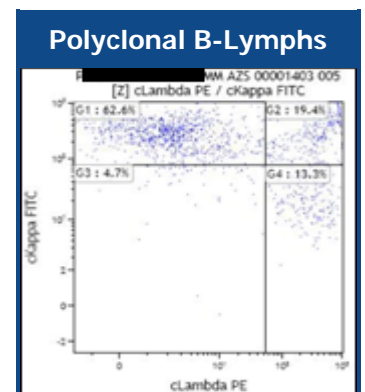
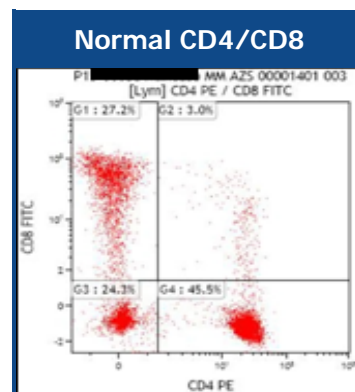
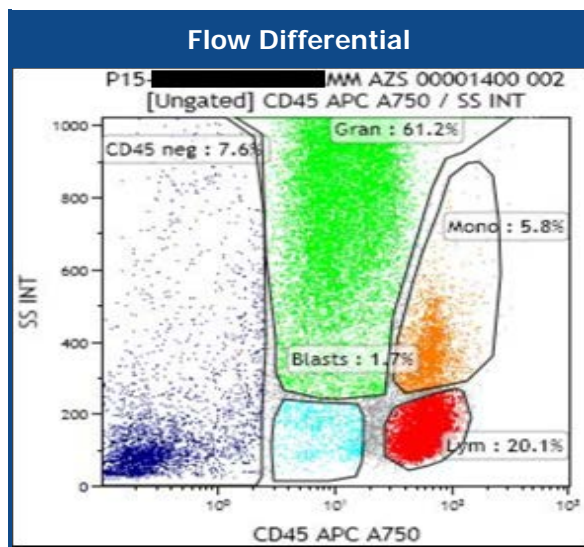
The lymphocytes (20%) include 17% B-cells with an excess of immunoglobulin kappa to lambda light chains (5:1), but are negative for CD5 and CD10. Seventy-four percent (74%) of the lymphocytes are mature T-cells with a normal CD4/CD8 ratio, and 10% natural killer (NK) cells. Less than 1% of the cellularity are plasma cells. Although excess cytoplasmic kappa light chain is detected, no cohesive population of cells is observed.

RESULT

Analysis Time: 4/12/19 13:04

Viability: 98% (Normal > 80%)

Specimen: BM, Lavender-top tube



Flow Cytometry Differential

Lymphocytes:	20%
Monocytes:	6%
Granulocytes:	61%
Plasma Cells:	<1%
Blasts:	2%
nRBC & Debris:	8%

Lymphocytes

Plasma Cells

Marker	%	Marker	%
CD2	82	CD19	93
CD3	74	CD56	17
CD4	48	cKappa	54
CD5	68	cLambda	4
CD7	81	IgA	19
CD8	25	IgG	47
CD10	2	IgM	87
CD19	17		
CD20	17		
CD38	13		
CD45	100		
CD56	10		
Kappa	13		
Lambda	6		

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DIAGNOSIS:

Bone marrow, aspirate: Normal female karyotype

INTERPRETATION

KARYOTYPE "ISCN": 46,XX[20]; Normal Female Karyotype

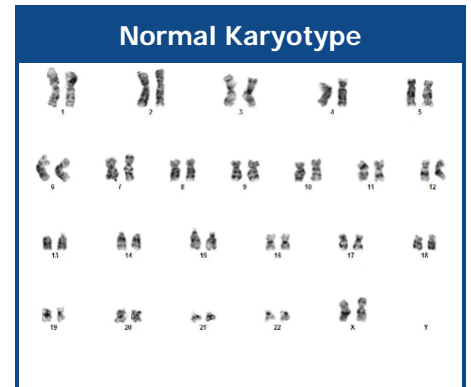
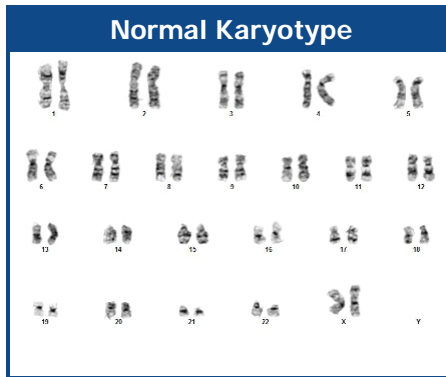
Conventional cytogenetic analysis shows a female karyotype with no evidence of an acquired clonal abnormality. This does not exclude the possibility of an abnormality that cannot be detected at the chromosomal level or exists at a low residual level.

Interpretation of this specimen's cytogenetic results should be made in conjunction with morphologic, immunophenotypic, and clinical findings. The results of this analysis do not exclude the possibility of genetic alterations below the band-resolution of this test or abnormalities due to other etiologies.

FISH studies are recommended and much more sensitive than G-band analysis for cases of plasma cell neoplasia because plasma cells have a very low proliferation rate in culture. FISH studies are recommended for clinically-significant abnormalities. (NCCN Guidelines, ver 2.2013, Multiple Myeloma, National Comprehensive Cancer Network, nccn.org)

Analysis

Cells Counted:	20
Cells Analyzed:	20
Cells Imaged:	3
Cells Karyotyped:	3
Band Level:	450
Banding Type:	G-Banding
Indication:	Monoclonal paraproteinemia



Electronically Signed By: Frank Bauer, MD (04/18/19 16:40)

DIAGNOSIS:

Bone marrow, aspirate: No Clonal Abnormalities Detected with probes specific for recurrent abnormalities in Plasma Cell Myeloma (see interpretation for probes tested).

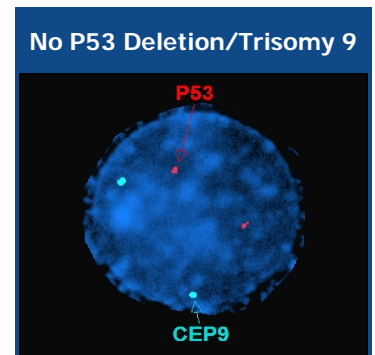
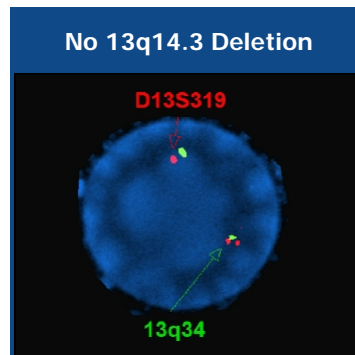
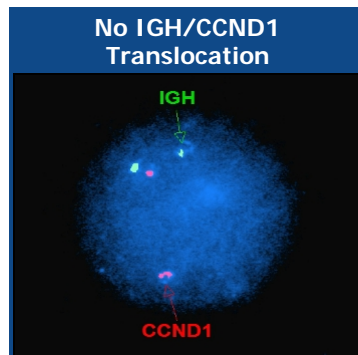
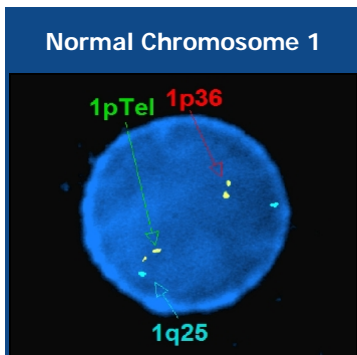
INTERPRETATION

FISH "ISCN": nuc ish (p58x2,1q25x2)[200],(CEP9x2)[200],(CCND1x2,IGHx2)[200],(D13S319x2,13q34x2)[200],(p53x2)[200]

Fluorescence in situ hybridization (FISH) with a panel of probes specific for detection of recurring chromosome abnormalities in plasma dysplasia was performed on uncultured bone marrow cells.

The regions/loci represented in these probe mixes were:

1. IGH/CCND1 dual color, dual translocation probes to 11q13 & 14q32 regions respectively, reveal a hybridization pattern within normal limits in 200 analyzed nuclei.
2. P53 (17p13.1), used to detect deletion/copy number abnormalities of chromosome 17p, reveals a hybridization pattern within normal limits in 200 analyzed nuclei.
3. 1p36 Microdeletion Region Probe - LSI p58 (1p36) /TelVysion 1p/LSI 1q25, used to detect copy number abnormalities of chromosome 1, reveal a hybridization pattern within normal limits in 200 analyzed nuclei.
4. CEP9 (centromere probe to chromosome 9), used to detect copy number abnormalities of chromosome 9, reveals a hybridization pattern within normal limits in 200 analyzed nuclei.
5. D13S319 (13q14.3) and 13q34, used to detect copy number abnormalities/deletion of chromosome 13, reveal a hybridization pattern within normal limits in 200 analyzed nuclei.



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DIAGNOSIS:

Bone marrow, aspirate: MYD88 mutation(S) detected: P.L265P (see comment)

INTERPRETATION

MYD88 mutation is the most frequent genomic abnormality in diffuse large cell lymphoma (DLBCL) activated B-cell-like (ABC) subtype, detected in 40% of cases. MYD88 is rarely mutated in the germinal center B-cell-like (GCB) DLBL, therefore, it can be used to differentiate between the two subtypes. MYD88 mutation is detected in approximately 90% of cases of Waldenstrom macroglobulinemia/lymphoplasmacytic lymphoma. MYD88 mutation analysis can be a useful prognostic tool for patients with IgM-MGUS since the L265P mutation is associated with a higher risk of disease progression and a greater disease burden. MYD88 mutation has also been reported to be common (40%) in central nervous system lymphoma.

METHODOLOGY:

Total nucleic acid was extracted from patient's plasma, PB/BM cells or paraffin-embedded tissues (FFPE). Bi-directional Sanger sequencing of exon 5 of MYD88 was performed, including the L265P mutation hot spot. This is a sequencing-based assay which has a typical sensitivity of 10-15% for detecting MYD88 mutations in a wild-type background. Various factors including quantity and quality of nucleic acid, sample preparation, and sample age can affect assay performance.

REFERENCES:

1. Trøen G, Warsame A, Delabie J. CD79B and MYD88 Mutations in Splenic Marginal Zone Lymphoma. *ISRN Oncol.* 2013;2013:252318.
2. Xu L, et al. MYD88 L265P in Waldenstrom's macroglobulinemia, IgM monoclonal gammopathy, and other B-cell lymphoproliferative disorders using conventional and quantitative allele-specific PCR. *Blood.* 2013;121(11):2051-2058.

The performance characteristics of this test have been determined by the laboratory. This test has not been approved by the FDA. The FDA has determined such clearance or approval is not necessary. The laboratory is CLIA certified to perform high complexity clinical testing.

Electronically Signed By: Frank Bauer, MD (04/18/19 16:36)

Patient: Jane A. Doe

Case #: X19-00352

Received Information: 2 Formalin containers, 10 smears, 2 green-top tubes, 1 lavender-top tube

Received: 04/09/19 11:04

Reported: 04/18/19 17:43